

Plasmid RD-PGA16

Quantity:	50 µg supercoiled plasmid DNA, ethanol-precipitated and air dried.
Storage and Reconstitution:	The dry pellet is shipped at room temperature. To reconstitute, spin the tube briefly and dissolve the contents in 50 µl of sterile, nuclease-free water to make a 1 µg/µl solution. Store at 4°C.

Note: A License from Molecular Devices Corporation is required prior to purchasing or using the plasmid.

Description

The cDNA for G α 16, the alpha-subunit of the promiscuous G-protein, was cloned from human hematopoietic cell line TF-1 by RT-PCR. When expressed in transfected cells, the G α 16 protein allows many Gi/o- and Gs-coupled receptors, which normally act through the cAMP pathway, to stimulate phospholipase C and mobilize intracellular calcium.

RD-PGA16 will be provided in two constructs with two different selectable markers (hygromycin, G418) to allow co-transfection with receptors.

pCEP-G α 16 (hygromycin)

- 1.5-kb human G α 16 cDNA in pCEP4 (vector size is 10.4-kb)
- Cytomegalovirus (CMV) enhancer/promoter for strong, constitutive expression of G α 16
- Hygromycin resistance gene for selection of stable transfected cells

RD-PGA16

- Epstein Barr Virus (EBV) origin of replication (oriP) and nuclear antigen (EBNA) for episomal replication in canine and primate cell lines
- Amp^r and ColE1 origin for selection and propagation in E. coli

pLEC1-G α 16 (G418)

- 1.5-kb G α 16 cDNA in pLEC1 (vector size is 5.5-kb)
- Cytomegalovirus (CMV) enhancer/promoter for strong, constitutive expression of G α 16
- Neomycin resistance gene for G418 selection of stable transfected cells
- T7 and SP6 promoters allowing *in vitro* synthesis of sense and antisense G α 16 RNA transcripts
- Amp^r and ColE1 origin for selection and propagation in E. coli

Quality Control

1. Identity and purity of the plasmid DNA is confirmed by restriction analysis on agarose gel.
2. Functional testing is carried out by transient transfection of the plasmid into CHO cells stably expressing the human nociceptin receptor (Gi/o-coupled) and measuring the stimulation of calcium mobilization by nociceptin in the FLIPRTM instrument (Molecular Devices Corporation, Sunnyvale, CA).



